COURSE DESCRIPTION (SYLLABUS)

	Course:
1.	Cell Culture Techniques
2.	Language of instruction:
	English
3.	Faculty:
	Faculty of Biotechnology
4.	Course/module code:
	29-BT-S1-E6-EnCCTc
5.	Course/module type (mandatory or elective):
	mandatory
6.	Programme:
	Biotechnology
7.	Study cycle (1st/2nd):
	1st cycle
8.	Year:
	3rd
9	Semester (autumn or spring):
9.	spring
10.	Form of tuition and number of hours:
	Laboratory: 30 h
11	Coordinator(s):
	Aleksandra Simiczyjew, PhD
12.	Initial requirements (knowledge, skills, social competences):
	Basic knowledge of biology and biochemistry at the level of first year undergraduate studies.
	Objectives:
13.	Students are going to gain basic knowledge about culturing animal cells. They will get elementary skills how to culture animal cells derived from normal tissues and cancer cell lines. General ken concerning fluorescence/confocal microscopy and its applications will be passed to the students. Students are also going to gain knowledge about quantitative methods used in the study of animal cells on the example of the assessment of the cell viability and cell cycle distribution under drug treatment.
14.	Content:
	Attending the module will give the students an opportunity to get familiar with speciality of working in the laboratory where animal cells are cultured. During the practical course students will learn how to isolate primary cells from chicken embryos

	and how to culture primary cells and tumour cell lines, how to count cells and passage them. Students will gai slides with fixed and stained cells and how to analyse distribution of selected actin cytoskeleton proteins wit inverted light microscope and confocal microscope, re- analysis like estimating cell viability and cell cycle distr done.	, how to examine cell viability, in also skills how to prepare cells morphology and thin a cell with the help of spectively. Some quantitative ribution in tested cells are also
	Learning outcomes:	Outcome symbols:
	Knowledge:	
	 Student has knowledge about basic techniques and research tools used in biochemistry, molecular biology and cell biology. 	K1_W08
	• Student knows the basic principles of safety rules and occupational health and ergonomics in the laboratory; student knows the rules of work in cell culture laboratory.	K1_W10
	Skills:	
	 Student applies basic physicochemical and biochemical techniques necessary for studying biological processes (including processes taking place in cultured cells). 	K1_U01
45	 Student has the skills in culturing cells of higher organisms 	K1_U02
15.	• Student carries out simple experiments under the guidance of a tutor in the field of animal cell biology, describes the results and present them to the tutor and other students.	K1_U05
	 Student uses appropriate scientific terminology in discussions with cell biology specialists. 	K1_U09
	 Student knows how to work as a part of team, work together to solve problems and perform scientific experiments. 	K1_U013
	Social competence:	
	 Student understand the need for continuing education throughout the whole life, including broadening knowledge in cell biology. 	K1_K01
	 Student understands the need for accurate planning of tasks and scientific experiments. 	K1_K03
	Recommended literature:	1
16.	• Culture of Animal cells - a manual of basic technic R.I. Freshney, Willey-Blackwell, 7 th edition, 2016.	ques and specialized applications,

17.	Methods of verification of the assumed learning outcomes:		
	written test		
18.	Conditions of earning credits:		
	active participation in laboratory classes		
	• passing the written test		
19.	Student's workload:		
	Activity	Number of hours for the activity	
	Hours of instruction (as stipulated in study programme):Lab.: 30 h	30 h	
	 Student's own work: preparation before classes: 5 h preparation for the test and final exam: 10 h 	15 h	
	Total number of hours:	45 h	
	Number of ECTS:	2 ECTS	